

Peri-implant Mucosal Tissues and Inflammation: Clinical Implications

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Purpose: The purpose of this review was to explore the available literature and compile studies that discuss the relevance of the biofilm, onset and progression of disease, critical peri-implant pocket depth, frequency of supportive implant therapy, excess cement, and keratinized peri-implant tissues as related to peri-implant disease. **Materials and Methods:** PubMed, Cochrane Oral Health Group Specialized Trial Register, and hand searches of related journals were performed in relationship to the focused question. Reports describing techniques, preclinical studies, and case reports were excluded. **Results:** Due to the absence of controlled studies, a meta-analysis could not be performed. Summaries of relevant publications were completed for each topic area. Clinical recommendations were developed to provide guidance to the practitioner. **Conclusion:** The importance of proper diagnosis, planning, and clinical treatment cannot be overstated. Patient factors including systemic disease, periodontal status, and oral hygiene significantly impact peri-implant health. Clinician factors such as implant position, excess cement, and restorative design can contribute to development of peri-implant disease. Surveillance of implant status is essential and can be assisted by the assessment of risk factors, establishment of a proper recall program, and monitoring changes in bone and peri-implant pocket depths. *INT J ORAL MAXILLOFAC IMPLANTS* 2019;34(suppl):s25–s33. doi: 10.11607/jomi.19suppl.g2

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Over the past almost five decades, dental implants have been a predictable therapy to replace missing teeth. The field has also evolved to include more

aggressive loading protocols, osseous augmentation of deficient bone areas, and modification of the implant designs. These, along with other factors, have led to an expansion in the number of implants placed and their more widespread use by both general practitioners and dental specialists. The importance of proper diagnosis, planning, and clinical treatment cannot be overstated. However, survival/success have been found to be affected by a number of risk factors resulting in peri-implant diseases. Peri-implant diseases have been characterized as a condition of the tissues around osseointegrated implants with signs of inflammation (bleeding and/or suppuration on probing) with or without loss of supporting bone.

Early on, Meffert (1992) described the ailing and failing implant.¹ The ailing implant had bone loss with pocketing but was static at maintenance visits, whereas the failing implant also demonstrated bone loss with pocketing but presented additionally with bleeding on probing, purulence, and continued bone loss despite therapy. Misch (1998) also utilized clinical parameters to assess implant health.² A continuum of health to disease was described with disease status related to a progressive worsening of clinical parameters

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such as probing depth, bone loss, and pain. Froum and Rosen (2012) established thresholds for clinical parameters as a basis for classification of peri-implantitis.³ Combinations of bleeding on probing and/or sup-puration, probing depth, and radiographic bone loss were utilized to classify peri-implantitis into early, moderate, and advanced categories. Recently, a classification system based on etiology was proposed.⁴ Its results indicated that the majority of bone loss was related to biofilm, followed by iatrogenic factors, exogenous irritants, absence of keratinized tissue, and extrinsic pathology.

Although the literature denotes many risk factors associated with peri-implant diseases, the purpose of this review was to explore the available literature and compile studies that discuss the relevance of the biofilm, onset and progression of disease, critical peri-implant pocket depth, frequency of supportive implant therapy, excess cement, and keratinized peri-implant tissues as related to peri-implant disease.

MATERIALS AND METHODS

Rationale

This systematic review evaluated the relationship of various risk factors as they pertain to dental implant disease.

Focused Question

In patients with dental implants, what is the relationship of biofilm, onset and progression of disease, critical peri-implant pocket depth, frequency of supportive implant therapy, excess cement, and keratinized peri-implant tissues with peri-implant disease?

Search Protocol

PubMed and Cochrane Oral Health Group Specialized Trial Register were searched. Hand searches were performed on *Clinical Oral Implants Research*, *The International Journal of Oral and Maxillofacial Implants*, *The International Journal of Periodontics and Restorative Dentistry*, *Journal of Clinical Periodontology*, *Journal of Periodontology*, and *Journal of Periodontal Research*.

Selection Criteria

Publications reporting mucositis of dental implants related to biofilm, keratinized peri-implant tissues, peri-implant pocket depth, frequency of recall, and residual cement were included in the analysis. Reports describing techniques were excluded.

Data Collection and Analysis

Due to the absence of controlled studies, a meta-analysis was not performed.

Biofilm and Inflammatory Response to Peri-implant Disease

The biofilm related to peri-implant diseases has not been extensively documented. The literature for peri-implant diseases follow those of gingivitis and periodontitis with regard to microflora and inflammatory response. As Loe et al (1965) induced an experimental gingivitis,⁵ several studies have described the response around dental implants.⁶⁻⁹ They indicate that the response to the cessation of oral hygiene around implants and its onset are similar to that around the natural teeth. The classic signs of inflammation—Gingival Index, bleeding on probing, and Plaque Index—were increased. Salvi et al (2012) also induced an experimental peri-implant mucositis and found similar results.⁶ However, the Gingival Index was higher around dental implants when compared to teeth. The authors postulated that the inflammatory response for dental implants is more severe than for the natural dentition. In addition, they found that with the reintroduction of oral hygiene measures, the soft tissues recovered to baseline values.⁶ Zitzmann et al (2001) evaluated the cellular response with an experimentally induced peri-implant mucositis with tissue biopsies.⁷ The response of T- and B-cells was similar to that around teeth. At 21 days, there was an increase in the volume of these lymphocytes. Lastly, host biomarkers including IL1- β , TNF- α , and TGF- β 2 have been evaluated in the induced peri-implant mucositis model.¹⁰ Only IL-1 β increased over the 3-week period in the gingival crevicular fluid. Following reinstatement of oral hygiene, IL-1 β levels returned to baseline. In natural longstanding peri-implant mucositis lesions, sites that bled on probing and had some degree of redness and inflammation demonstrated on biopsy infiltrate in the connective tissue.¹¹

The biofilm conversion from peri-implant mucositis to peri-implantitis has been only documented with cross-sectional studies describing the characteristics of peri-implantitis. The microbiome of healthy and/or diseased implants have been compared.^{12,13} In general, species of the red complex such as *Porphyromonas gingivalis* and *Tannerella forsythia* have been found in diseased sites. Sanz-Martin et al (2017) studied healthy and diseased implants, illustrating that both states had a core microbiome, while health had taxa consistent with periodontal health and peri-implantitis had taxa consistent with periodontitis.¹⁴ In addition, the findings regarding peri-implantitis biofilm have shown species including *Synergistetes* and *Tannerella*. In a systematic review by Lafaurie et al (2017), peri-implantitis was an infection with a diversity of microorganisms including periodontal pathogens, anaerobic Gram-negative rods, and rarely, enteric rods and *Staphylococcus aureus*.¹⁵ Overall, the majority of information

available indicates that a shift occurs from health to disease with regard to the biofilm. The biofilm associated with the diseased dental implant seems to be more pathogenic and somewhat like those in periodontitis.

Clinical Recommendations

- The clinician should be aware that a shift in the biofilm and inflammatory response occurs with the conversion of a healthy to diseased implant state. For example, with the deepening of a peri-implant pocket, the practitioner should consider additional measures to monitor and/or treat the failing implant.
- The biofilm associated with peri-implant disease may be similar to that in periodontitis. As a result, the susceptible periodontitis patient with dental implants should be carefully monitored.
- Given the inflammatory basis of peri-implant diseases, the surgical and restorative clinician should carefully consider the contour, function, and overall cleansability of the prosthesis throughout the planning process.

Time to Onset and Progression of Peri-implant Diseases

The development of peri-implant diseases seems dependent on many variables, such as oral hygiene, quality of tissue, and restoration type. However, unlike periodontitis, factors such as position of the implant within the edentulous ridge, implant surface texture, etc, impact the development of both mucositis and/or peri-implantitis. In addition, the pattern and rate of onset do not follow that of periodontal disease but may be influenced by the presence or history of periodontitis.

Studies involving experimentally induced peri-implantitis demonstrate clinical changes as in an induced gingivitis.⁶ The cessation of oral hygiene produced increases in plaque, Gingival Index, and bleeding. When oral hygiene was reinstated, clinical parameters recovered.⁸ The analysis of biologic responses to the induction of peri-implant mucositis indicates an upregulation of peri-implant crevicular fluid factors and the cellular response involving an increase in the proportions of T and B cells.⁷ Overall, these studies indicate that the clinical biologic response as well as timing of progression in the experimental induction of peri-implant mucositis collates with induced gingivitis.

In a retrospective review of dental radiographs, Fransson et al (2010) evaluated the onset and progression of bone loss.¹⁶ A total of 182 patients with 419 implants were assessed for bone loss and had a mean follow-up period of 11.1 years. Bone loss after the first year averaged 1.68 mm. Interestingly, 32% of the implants had bone loss of ≥ 2 mm, and 10% of those had bone loss ≥ 3 mm. Analysis indicated that progression

was nonlinear and the rate decreased with time. Derks et al (2016) also found that the pattern of bone loss was nonlinear.¹⁷ A retrospective review of 105 implant dental radiographs indicated that the onset generally occurred within the first 3 years of function. However, compared to Fransson et al (2010),¹⁶ the amount of bone loss was greater with an average of 3.5 mm. Bone loss of more than 3 mm occurred in 51% of the sample. The authors postulated that the bone loss could be related to a prior history of periodontitis. In a shorter-duration study, Schwarz et al (2017) found that of 512 implants, a total of 262 were diagnosed with peri-implant disease.¹⁸ The majority of disease that occurred with implants was found between 12 and 48 months. After 48 months, the rate of disease was similar to that between 1 and 12 months. These results again indicate that peri-implant disease occurs early in the life of a functionally loaded implant. Recently, Sarmiento et al (2018) found that the timing of grafting influenced the rate of peri-implant disease.¹⁹ In a staged grafting procedure where the implant is placed following a healing period, the rate of peri-implant mucositis was 8.1% and the rate of peri-implantitis was 4.4%. However, when grafting occurred at the time of implant placement, the rate of peri-implant mucositis was 7.5% and the rate of peri-implantitis was 9.7%. In addition, the onset of mucositis preceded the statistical detection of radiographic bone loss.

Clinical Recommendations

- Although limited, studies indicate that the onset and the majority of peri-implant disease progressions occur in the life of a functionally loaded implant generally prior to 36 months.
- Since the onset and progression of peri-implant diseases seem not to follow patterns of periodontitis, the clinician should have an enhanced awareness of risk and should structure an individualized recall program for the patient.

Critical Peri-implant Pocket Depth

The establishment of the biologic width with implant healing has been well documented. Although the dimensions seem to vary when compared to the natural dentition, the formation of an epithelial and connective tissue attachment occurs. The healing results in the formation of a peri-implant sulcus. Several factors, such as surface characteristics, implant height above the bone level, platform switching, microgap, abutment size, etc, influence the final measurements. With the onset of peri-implant disease, these structures begin to change much in the same manner as with periodontitis. Gingival inflammation can increase the peri-implant sulcus and with loss of supporting bone, apical migration of the connective tissue and

epithelium occurs. The disease around an implant with bone loss then forms a deepened peri-implant pocket.

Healthy peri-implant mucosa can be altered by factors that progress to conditions of peri-implant mucositis and peri-implantitis. Although clinical assessment can differentiate between a state of health and disease, a critical implant pocket depth can assist in defining this transitional state. Derks et al (2016) described healthy peri-implant tissue by the absence of bleeding on probing, mucositis as presence of bleeding on probing \pm suppuration without radiographic bone loss, and peri-implantitis as bleeding on probing \pm suppuration and radiographic bone loss > 0.5 mm (45% of cases presented).¹⁷ The onset of disease occurred early, with the majority of peri-implantitis cases/implants occurring within 3 years of function. Schwarz et al (2018) measured peri-implant sulcus/pocket depth values of 1 to 3 mm and 4 to 6 mm for healthy and mucositis implants, respectively.²⁰ Peri-implantitis implants only recorded pocket values in the 4- to 6-mm range. Schwarz et al (2017) noted that mucositis and peri-implantitis could present within 12 to 48 months after initial implant placement.¹⁸

Recently, Monje et al (2018) evaluated 1,572 sites around 262 implants in 141 patients.²¹ The clinical parameters included the evaluation of pocket depth. In this cross-sectional matched case-control study, healthy implants had a mean pocket depth of 2.63 mm, mucositis-classified implants had a mean pocket depth of 3.26 mm, and peri-implantitis–diagnosed implants had a mean pocket depth of 4.58 mm. The authors concluded that pocket depth might accurately discern between diagnoses among peri-implant conditions. Similarly, Ramanauskaite et al (2018) evaluated 269 dental implants with several different clinical parameters including peri-implant pocket depth.²² A total of 77 dental implants diagnosed as healthy had a mean pocket depth of 2.95 mm. Peri-implant mucositis implants ($n = 77$) had a mean pocket depth of 3.10 mm, whereas peri-implantitis implants ($n = 115$) had a mean pocket depth of 4.91 mm. These differences in pocket depth between the three groups were significantly different.

Clinical Recommendations

- The documentation of dental implant health status is an important component of clinical care. During the treatment process, a baseline radiograph (at final prosthesis insertion) and clinical parameters (pocket depth, bleeding on probing, etc) should be recorded. The frequency of periodic documentation should be based on individual risk factors in order to assess changes in implant health over the life of the implant.

- The apparently healthy implant may still harbor an environment (biofilm, inflammation, etc) that could predispose the implant to convert to disease. Studies indicate that the clinician should be aware of the development of future markers for peri-implant disease and conditions including profuse bleeding on probing, suppuration, bone loss, and critical implant pocket depth of ≥ 5 mm. This awareness includes but is not limited to continuing routine care, radiographic assessment, oral hygiene instruction, and shortened recall interval.

Frequency of Supportive Implant Therapy

According to contemporary studies, there is an estimated 43% prevalence of peri-implant mucositis and a 22% prevalence of peri-implantitis.^{23–25} A majority of this peri-implant disease may share a pathogenesis with periodontitis.²⁶ Therefore, peri-implant maintenance seems to be critical to maintain the stability of the tissue around dental implants.^{27,28}

Despite the understanding of the importance of peri-implant maintenance, there have been no well-controlled investigations that refine the peri-implant recall interval; this is due to various definitions of diagnosis, unclear terminology, varying follow-up periods, and ethical considerations. Thus, most of the evidence of supportive implant therapy interval has been obtained from retrospective studies or reviews.

The prevalence rates of peri-implant diseases were evaluated in 89 patients.²⁹ The patients were assigned to 3-month intervals during the first year after implant placement and later on 6-month intervals. Patients who did not participate in regular supportive implant therapy had an 11-fold higher chance of peri-implantitis than patients showing good compliance. On the contrary, patients who did not have regular supportive implant therapy were reported to have up to 48% of the prevalence of peri-implant mucositis during an observation period of 9 to 14 years.³⁰ These studies have confirmed the essential role of supportive implant therapy to maintain tissue health and that the lack of supportive implant therapy will lead to a higher prevalence of the peri-implant disease.

A retrospective study done by Frisch et al (2015) evaluated the compliance and the frequency of the supportive post-implant therapy program and indicated a positive correlation between lower compliance and increased probing depth and higher plaque rate.³¹ The compliance rates were categorized into five different groups, from a 3-month interval to no compliance at all. The higher rates of patient compliance (86% to 94%) were observed during the first 3 years. A significant correlation was found between lower compliance and increased peri-implant probing depth. In addition,

higher plaque rates were found in individuals with lower compliance rates.

Dental implant patients may have a higher risk of peri-implant disease due to diabetes, poor oral hygiene, and smoking.^{29,32-34} A history of periodontitis seems to be the most documented risk for future peri-implant disease.^{28,35,36} Long-term studies done by Rocuzzo and coworkers evaluated the tissue around dental implants with supportive implant therapy for 10 years.³⁷⁻³⁹ The study found that in periodontally healthy patients there were no statistical differences in clinical parameters if subjects adhered to their supportive implant therapy or not. In contrast, the patients who had moderate to severe periodontal disease had higher plaque scores, bleeding scores during the supportive implant therapy, and eventually implant loss. In a systematic review, Monje et al (2016) analyzed 13 studies to evaluate the impact of supportive maintenance on the implant.⁴⁰ This review successfully provides positive evidence of the patient with maintenance and concluded a minimum recall post-implant maintenance therapy of 5 to 6 months. However, the studies did indicate that the possibility of biologic complications might still occur and other risk factors should be appropriately considered.

In 2013, Aguirre-Zorzano et al stated that the prevalence of peri-implant inflammatory disease in periodontal patients who regularly undergo supportive implant therapy with a mean recall of 4 months is clinically significantly lower and the peri-implant disease could be even prevented.⁴¹ A similar result was found in a retrospective study by Costa et al (2012), which suggested that the simple fact of enrolling subjects for supportive implant therapy may reduce the risk of peri-implantitis from 43.9% to 18% at the patient level with maintenance at least once a year.⁴² These findings provide clinical evidence that the supportive implant therapy interval should be adjusted case by case, particularly in patients with a history of periodontitis.

Clinical Recommendations

- In general, a reasonable interval of supportive implant therapy is 5 to 6 months for a patient with low risk of peri-implant disease, but should be evaluated case by case.
- When considering a patient with a history of risk, such as periodontal disease, it may be necessary to shorten the supportive implant therapy interval.
- During a supportive implant therapy appointment, reinforcement of oral hygiene, modification of factors such as smoking, and removal and cleaning of the prosthesis should be considered.

Role of Excess Cement in Peri-implant Diseases

Cemented restorations are commonly used to restore dental implants. It has been shown that excess cement left in the sulcus around implant-supported restorations can cause inflammation, ultimately leading to peri-implant disease. Cemented restorations offer several advantages, such as ease of prosthetic fabrication, reduced costs, increased framework passivity, and improved esthetics due to the absence of the buccal screw access hole. The main disadvantage of cement-retained restorations is the fact that cements are a flowable material that can spread unintentionally to the adjacent gingival tissues, making it difficult for the clinician to properly remove excess subgingival material. Therefore, caution on implant selection, placement, and prosthetic design should play a key role in the initial treatment plan. Although we aim for screw restoration as a primary election to restore a dental implant, it is not always feasible if angulation of the implant body is necessary.

A study published by Wilson (2009) demonstrated how the presence of peri-implantitis caused by dental cements was observed in 81% of cases.⁴³ Korsch and Walther (2015) compared various types of cements and associated peri-implant disease. The authors found that the frequency of excess cement depended on cement type.⁴⁴ For implants that used methacrylate cement, the frequency of excess cement was 62%, whereas the frequency of excess cement for zinc oxide-eugenol cement was 100%. Although several publications have documented different peri-implant complications, few have addressed the causative factor for the biologic breakdown. In a study published by Sarmiento et al (2016), the classification system based on etiologies included 5.5% of cases with peri-implantitis induced by an exogenous factor, eg, residual excess cement.⁴ Another retrospective study by Linkevicius et al (2013) concluded that patients with previous periodontal disease may also be more predisposed to peri-implantitis due to excess cement.⁴⁵ Other studies have shown that the presence of suppuration around dental implants was greater on crowns that were cemented versus screw-retained.⁴⁶ Many authors have shown different techniques to reduce excess cement complications. Linkevicius et al (2013) in a clinical study showed that dental radiographs should not be considered as a reliable method for cement excess evaluation.⁴⁷ His results revealed that cement remnants were around 7.5% to 11.5% depending on the location of detection. Wadhvani et al (2010) proved that radiographic density of implant restorative cements is poor and depending

on the thickness of the cement, smaller pieces would remain unseen.⁴⁸ They suggested placing margins supragingival for proper cement removal.

Challenges for the clinician on when to place a cement-retained restoration should not be based on material selection to achieve higher esthetics, but rather the criteria should be those unique to cementation technique and proper treatment planning. Therefore, avoiding cement-retained restorations in difficult-to-access areas should be recommended for clinicians with less experience.

Clinical Recommendations

- All attempts to reduce excess submarginal cement should be made through the use of screw-retained restorations or abutments designed to minimize the depth of the cement line in relation to the peri-implant tissue architecture. The cement line should be located with a minimal depth for ease of removal and to minimize submarginal excess cement.
- A final radiograph should be carried out routinely to assess unwanted cement beyond the gingival margin.
- Postoperative assessment is recommended within the first weeks post-crown insertion. This should allow immediate clinical detection of excess cement and avoid peri-implant tissue inflammation.

Role of Keratinized Tissue Surrounding Dental Implants

Peri-implant health has been defined both clinically and histologically.⁴⁹ Peri-implant tissues surround an osseointegrated dental implant and can be divided into hard and soft tissue components. The hard tissue component forms a contact relationship to the implant surface, which contributes to implant stability.⁵⁰ The soft tissue component is formed during the healing process following implant/prosthetic placement.⁵¹ The peri-implant tissues protect the bone that supports the implant. With the absence of healthy peri-implant tissues, the long-term implant success and survival becomes compromised and less predictable.⁴² However, if there is an insufficient amount of keratinized tissue around implants, is there an increased risk of peri-implant mucositis? If so, what types of modalities are appropriate to address a lack of keratinized tissue of the peri-implant tissues?

Does the Absence of Keratinized Tissue Influence Peri-implant Mucositis? Keratinized tissue or mucosa, which extends from the margin of the peri-implant mucosa to the mucogingival junction, is composed of fibrous connective tissue with fibroblasts; type I, III, IV, V, and VI collagen; and an orthokeratinized squamous epithelium.^{52,53} Keratinized gingiva has been defined

as marginal and attached gingiva that excludes soft tissue of the interdental col region—interproximal gingival tissue between posterior teeth where epithelium is devoid of keratinization.⁵⁴ One theory explaining the reduction of keratinized tissue is the post-tooth extraction natural loss of crestal bone. The buccal thickness of keratinized tissue is greater at the base of implants than at teeth (2.0 mm vs 1.1 mm, respectively).⁵³ According to a new classification scheme for periodontal and peri-implant diseases and conditions, peri-implant mucositis is defined as an inflammatory lesion of the mucosa surrounding an endosseous implant without loss of supporting peri-implant bone.^{34,55,56} This is clinically determined by the presence of redness, swelling, bleeding on probing, and suppuration. The dimensions of peri-implant keratinized mucosa may be a risk indicator for peri-implant mucositis. The need for a minimum amount of keratinized tissue in order to maintain peri-implant tissue health has been a controversial issue.^{57–61} Some studies suggested that plaque accumulation that resulted in marginal inflammation was more frequent at implant sites with < 2 mm of keratinized tissue.^{62–66} However, several studies suggested that the lack of a minimum amount of keratinized tissue was not associated with mucosal inflammation.^{64,67–72}

A systematic review assessed seven cross-sectional and four longitudinal studies to determine if keratinized mucosa affected implant health, suggesting that a lack of adequate keratinized tissue around endosseous dental implants is associated with plaque accumulation, tissue inflammation, recession, and attachment loss.⁵⁸ This supported a meta-analysis that indicated a statistically significant difference between plaque scores and modified Gingival Index in favor of sites with a wider dimension for keratinized tissue.⁵⁹ The width of keratinized tissue at implant sites is another area of controversy.⁷³ The association of keratinized mucosa width at implant sites was studied in a small group of patients 5 to 10 years retrospectively. Statistical analysis failed to indicate an association between keratinized tissue or the mobility of marginal mucosa around implant sites with plaque accumulation, bleeding on probing, or probing depth.⁶⁹ Another longitudinal study of 339 implants measuring the amount of keratinized mucosa present over a period of at least 3 years showed a higher Gingival Index (0.9 vs 0.8) and modified Plaque Index (1.5 vs 1.3) in patients with keratinized tissue with < 2 mm and > 2 mm, respectively.⁶² A 5-year study involving 307 implants in edentulous mandibles with fixed implant-retained reconstructions assessed sites with < 2 mm and > 2 mm of keratinized tissue. The investigators reported higher plaque scores (0.7 vs 0.4) and bleeding on probing (0.2 vs 0.1) at lingual sites as well as

recession (0.7 vs 0.1) at buccal sites.⁷² Another study involving 15 patients with mandibular overdentures on four implants assessed the presence or absence of keratinized tissue on buccal aspects of implants, showing that 19 implants with at least 2 mm of keratinized mucosa had lower plaque (0.3 vs 0.6) and gingival indices (0.1 vs 0.6) than 17 implants without keratinized mucosa.⁶⁶ It has been suggested that the evidence is equivocal regarding the effect of keratinized tissue on the long-term health of the peri-implant tissue. The notable advantages are the ease of plaque removal and patient comfort.⁷⁴⁻⁷⁸

Does Grafting Help in the Management of Peri-implant Mucositis? Historically, scientific evidence reported that a lack of keratinized tissue was not critical to maintain peri-implant soft tissue health nor to result in more peri-implant diseases.^{69,79} However, based on current evidence, it has been suggested that an increased amount of keratinized tissue may better preserve both soft and hard tissue stability, resulting in a favorable long-term outcome for dental implants (as well as better oral hygiene maintenance over time).^{63,72,73} Also, an increased amount of keratinized tissue thickness may decrease the risk of recessions with immediate implants.⁸⁰ Therefore, periodontal surgical procedures that augment soft tissue volume are recommended for esthetic and dimensional advantages following tooth extraction and implant therapy for both immediate and delayed placement.⁸¹⁻⁸⁴

Bleeding on probing was discussed in two studies with respect to grafting and nongrafting treatments after implant placement. According to a long-term study by Rocuzzo et al, there was an insignificant difference (23% and 27%) between groups with or without soft tissue grafting.⁶⁵ However, in another study there was notable improvement from mean baseline values of 85% to 30% with autogenous soft tissue grafting compared with 40% to 95% to 25% to 95% without soft tissue grafting.⁸³ In the same study, the mean Gingival Index shared a similar notable improvement comparing soft tissue grafting with no soft tissue grafting after follow-up periods of 6 to 12 months. Multiple studies indicated that there was a significant benefit to lower plaque values following surgical intervention when increasing keratinized tissue.⁶⁵ Conversely, one study compared Plaque Index of treated and untreated groups over time and found no significant difference at baseline and at 12 months.⁸⁵

With regard to probing depth, there were no significant changes over time between the different treatment groups of apically positioned flap versus apically positioned flap plus free gingival graft in a meta-analysis.⁸⁶ The mean probing depth values at baseline for soft tissue grafting, 1.97 to 3.09 mm, reduced to 2.08 to 3.18 mm after 6 to 12 months, while no soft tissue

grafting ranged from 1.76 to 3.25 mm at baseline and 1.60 to 3.62 mm after 6 to 12 months. Comparing these final probing depth values favored the apically positioned flap plus autogenous tissue. In a clinical study of 30 patients with keratinized tissue of < 1 mm at implant sites, half of the patients underwent surgery to widen the band of keratinized mucosa. After 10 years, there was a significant difference in the gain of keratinized mucosa of 3.1 mm versus 0 mm in patients who underwent surgery and patients who did not, respectively. However, the Plaque Index, bleeding on probing, and presence of peri-implantitis was not different between both groups.³¹ Lastly, a meta-analysis assessing three different treatment modalities (autogenous, collagen matrix, and apically positioned flap) used for implant site maintenance with > 2 mm, < 2 mm, or no keratinized tissue resulted in statistically significant differences favoring apically positioned flap plus autogenous tissue⁶⁵ in a period of 6 months.

Although there has been an increase of literature to support the use of soft tissue augmentation for keratinized tissue around peri-implant tissues, unfortunately there is a lack of data regarding clinical long-term outcomes associated with peri-implant soft tissue augmentation procedures.

Clinical Recommendations

- The removal of plaque around peri-implant tissues should be performed as part of a routine periodontal maintenance program to prevent the progression of peri-implant diseases and conditions.
- Soft tissue augmentation surgery should be performed around dental implants to provide stable keratinized tissue.

CONCLUSIONS

The importance of proper diagnosis, planning, and clinical treatment cannot be overstated. Patient factors including systemic disease, periodontal status, and oral hygiene significantly impact peri-implant health. Clinician factors such as implant position, excess cement, and restorative design can contribute to development of peri-implant disease. Surveillance of implant status is essential and can be assisted by assessment of risk factors, establishment of a proper recall program, and monitoring changes in bone and peri-implant pocket depths.

DISCLAIMER

The authors have no direct financial interests with the products and instruments listed in the paper.

REFERENCES

1. Meffert RM. Treatment of the ailing, failing implant. *J Calif Dent Assoc* 1992;20:42–45.
2. Misch CE. The implant quality scale: A clinical assessment of the health-disease continuum. *Oral Health* 1998;88:15–20, 23–25; quiz 25–26.
3. Froum SJ, Rosen PS. A proposed classification for peri-implantitis. *Int J Periodontics Restorative Dent* 2012;32:533–540.
4. Sarmiento HL, Norton MR, Fiorellini JP. A classification system for peri-implant diseases and conditions. *Int J Periodontics Restorative Dent* 2016;36:699–705.
5. Loe H, Theilade E, Jensen SB. Experimental gingivitis in man. *J Periodontol* 1965;36:177–187.
6. Salvi GE, Aglietta M, Eick S, Sculean A, Lang NP, Ramseier CA. Reversibility of experimental peri-implant mucositis compared with experimental gingivitis in humans. *Clin Oral Implants Res* 2012;23:182–190.
7. Zitzmann NU, Berglundh T, Marinello CP, Lindhe J. Experimental peri-implant mucositis in man. *J Clin Periodontol* 2001;28:517–523.
8. Pontoriero R, Tonelli MP, Carnevale G, Mombelli A, Nyman SR, Lang NP. Experimentally induced peri-implant mucositis. A clinical study in humans. *Clin Oral Implants Res* 1994;5:254–259.
9. Meyer S, Giannopoulou C, Courvoisier D, Schimmel M, Müller F, Mombelli A. Experimental mucositis and experimental gingivitis in persons aged 70 or over. Clinical and biological responses. *Clin Oral Implants Res* 2017;28:1005–1012.
10. Schierano G, Pejrone G, Brusco P, et al. TNF-alpha TGF-beta2 and IL-1beta levels in gingival and peri-implant crevicular fluid before and after de novo plaque accumulation. *J Clin Periodontol* 2008;35:532–538.
11. Gualini F, Berglundh T. Immunohistochemical characteristics of inflammatory lesions at implants. *J Clin Periodontol* 2003;30:14–18.
12. Carcuac O, Derks J, Charalampakis G, Abrahamsson I, Wennström J, Berglundh T. Adjunctive systemic and local antimicrobial therapy in the surgical treatment of peri-implantitis: A randomized controlled clinical trial. *J Dent Res* 2016;95:50–57.
13. Mombelli A, Décaillot F. The characteristics of biofilms in peri-implant disease. *J Clin Periodontol* 2011;38(suppl 11):203–213.
14. Sanz-Martin I, Doolittle-Hall J, Teles RP, et al. Exploring the microbiome of healthy and diseased peri-implant sites using Illumina sequencing. *J Clin Periodontol* 2017;44:1274–1284.
15. Lafaurie GI, Sabogal MA, Castillo DM, et al. Microbiome and microbial biofilm profiles of peri-implantitis: A systematic review. *J Periodontol* 2017;88:1066–1089.
16. Fransson C, Tomasi C, Pikner SS, et al. Severity and pattern of peri-implantitis-associated bone loss. *J Clin Periodontol* 2010;37:442–448.
17. Derks J, Shaller D, Håkansson J, Wennström JL, Tomasi C, Berglundh T. Effectiveness of implant therapy analyzed in a Swedish population: Prevalence of peri-implantitis. *J Dent Res* 2016;95:43–49.
18. Schwarz F, Becker K, Sahn N, Horstkemper T, Rousi K, Becker J. The prevalence of peri-implant disease for two-piece implants with an internal tube-in-tube connection: A cross sectional analysis for 512 implants. *Clin Oral Implants Res* 2017;28:24–28.
19. Sarmiento H, Norton M, Luan K, et al. The relationship between grafted sites and peri-implant disease. Scientific poster presented at the first US National Osteology Symposium, February 9, 2018, Phoenix, Arizona.
20. Schwarz F, Derks J, Monje A, Wang HL. Peri-implantitis. *J Periodontol* 2018;89(suppl 1):s267–s290.
21. Monje A, Caballé-Serrano J, Nart J, Peñarocha D, Wang HL, Rakic M. Diagnostic accuracy of clinical parameters to monitor peri-implant conditions: A matched case-control study. *J Periodontol* 2018;89:407–417.
22. Ramanauskaitė A, Becker K, Schwarz F. Clinical characteristics of peri-implant mucositis and peri-implantitis. *Clin Oral Implants Res* 2018;29:551–556.
23. Tonetti MS, Chapple IL, Jepsen S, Sanz M. Primary and secondary prevention of periodontal and peri-implant diseases: Introduction to, and objectives of the 11th European Workshop on Periodontology consensus conference. *J Clin Periodontol* 2015;42(suppl 16):s1–s4.
24. Atieh MA, Alsabeeha NH, Faggion CM Jr, Duncan WJ. The frequency of peri-implant diseases: A systematic review and meta-analysis. *J Periodontol* 2013;84:1586–1598.
25. Derks J, Tomasi C. Peri-implant health and disease: A systematic review of current epidemiology. *J Clin Periodontol* 2015;42(suppl 16):s158–s171.
26. Lang NP, Berglundh T; Working Group 4 of Seventh European Workshop on Periodontology. Periimplant diseases: Where are we now?—Consensus of the Seventh European Workshop on Periodontology. *J Clin Periodontol* 2011;38(suppl 11):178–181.
27. Jepsen S, Berglundh T, Genco R, et al. Primary prevention of peri-implantitis: Managing peri-implant mucositis. *J Clin Periodontol* 2015;42(suppl 16):s152–s157.
28. Pjetursson BE, Helbling C, Weber HP, et al. Peri-implantitis susceptibility as it relates to periodontal therapy and supportive care. *Clin Oral Implants Res* 2012;23:888–894, erratum 1004.
29. Rinke S, Ohl S, Ziebolz D, Lange K, Eickholz P. Prevalence of periimplant disease in partially edentulous patients: A practice-based cross-sectional study. *Clin Oral Implants Res* 2011;22:826–833.
30. Roos-Jansåker AM. Long time follow up of implant therapy and treatment of peri-implantitis. *Swed Dent J Suppl* 2007:7–66.
31. Frisch E, Ziebolz D, Vach K, Ratka-Krüger P. The effect of keratinized mucosa width on peri-implant outcome under supportive postimplant therapy. *Clin Implant Dent Relat Res* 2015;17(suppl 1):e236–e244.
32. Kotsovilis S, Karoussis IK, Fourmousis I. A comprehensive and critical review of dental implant placement in diabetic animals and patients. *Clin Oral Implants Res* 2006;17:587–599.
33. Serino G, Ström C. Peri-implantitis in partially edentulous patients: Association with inadequate plaque control. *Clin Oral Implants Res* 2009;20:169–174.
34. Lindhe J, Meyle J; Group D of European Workshop on Periodontology. Peri-implant diseases: Consensus Report of the Sixth European Workshop on Periodontology. *J Clin Periodontol* 2008;35(suppl 8):282–285.
35. De Boever AL, De Boever JA. Early colonization of non-submerged dental implants in patients with a history of advanced aggressive periodontitis. *Clin Oral Implants Res* 2006;17:8–17.
36. Konstantinidis IK, Kotsakis GA, Gerdes S, Walter MH. Cross-sectional study on the prevalence and risk indicators of peri-implant diseases. *Eur J Oral Implantol* 2015;8:75–88.
37. Rocuzzo M, De Angelis N, Bonino L, Aglietta M. Ten-year results of a three-arm prospective cohort study on implants in periodontally compromised patients. Part 1: Implant loss and radiographic bone loss. *Clin Oral Implants Res* 2010;21:490–496.
38. Rocuzzo M, Bonino F, Aglietta M, Dalmaso P. Ten-year results of a three arms prospective cohort study on implants in periodontally compromised patients. Part 2: Clinical results. *Clin Oral Implants Res* 2012;23:389–395.
39. Rocuzzo M, Bonino L, Dalmaso P, Aglietta M. Long-term results of a three arms prospective cohort study on implants in periodontally compromised patients: 10-year data around sandblasted and acid-etched (SLA) surface. *Clin Oral Implants Res* 2014;25:1105–1112.
40. Monje A, Aranda L, Diaz KT, et al. Impact of maintenance therapy for the prevention of peri-implant diseases: A systematic review and meta-analysis. *J Dent Res* 2016;95:372–379.
41. Aguirre-Zorzano LA, Vallejo-Aisa FJ, Estefanía-Fresco R. Supportive periodontal therapy and periodontal biotype as prognostic factors in implants placed in patients with a history of periodontitis. *Med Oral Patol Oral Cir Bucal* 2013;18:786–792.
42. Costa FO, Takenaka-Martinez S, Cota LO, Ferreira SD, Silva GL, Costa JE. Peri-implant disease in subjects with and without preventive maintenance: A 5-year follow-up. *J Clin Periodontol* 2012;39:173–181.
43. Wilson TG Jr. The positive relationship between excess cement and peri-implant disease: A prospective clinical endoscopic study. *J Periodontol* 2009;80:1388–1392.
44. Korsch M, Walther W. Peri-implantitis associated with type of cement: A retrospective analysis of different types of cement and their clinical correlation to the peri-implant tissue. *Clin Implant Dent Relat Res* 2015;17(suppl 2):e434–e443.

45. Linkevicius T, Puisys A, Vindasiute E, Linkeviciene L, Apse P. Does residual cement around implant-supported restorations cause peri-implant disease? A retrospective case analysis. *Clin Oral Implants Res* 2013;24:1179–1184.
46. Wittneben JG, Joda T, Weber HP, Brägger U. Screw retained vs. cement retained implant-supported fixed dental prosthesis. *Periodontol* 2000 2017;73:141–151.
47. Linkevicius T, Vindasiute E, Puisys A, Linkeviciene L, Maslova N, Puriene A. The influence of the cementation margin position on the amount of undetected cement. A prospective clinical study. *Clin Oral Implants Res* 2013;24:71–76.
48. Wadhvani C, Hess T, Faber T, Piñeyro A, Chen CS. A descriptive study of the radiographic density of implant restorative cements. *J Prosthet Dent* 2010;103:295–302.
49. Araujo MG, Lindhe J. Peri-implant health. *J Periodontol* 2018;89(suppl 1):s249–s256.
50. Albrektsson T, Sennerby L. State of the art in oral implants. *J Clin Periodontol* 1991;18:474–481.
51. Tomasi C, Tessarolo F, Caola I, Wennström J, Nollo G, Berglundh T. Morphogenesis of peri-implant mucosa revisited: An experimental study in humans. *Clin Oral Implants Res* 2014;25:997–1003.
52. Romanos GE, Schröter-Kermani C, Weingart D, Strub JR. Healthy human periodontal versus peri-implant gingival tissues: An immunohistochemical differentiation of the extracellular matrix. *Int J Oral Maxillofac Implants* 1995;10:750–758.
53. Chang M, Wennström JL, Odman P, Andersson B. Implant supported single-tooth replacements compared to contralateral natural teeth. Crown and soft tissue dimensions. *Clin Oral Implants Res* 1999;10:185–194.
54. Laney WR. Glossary of oral and maxillofacial implants. *Int J Oral Maxillofac Implants* 2017;32:Gi–G200.
55. Heitz-Mayfield LJA, Salvi GE. Peri-implant mucositis. *J Periodontol* 2018;89(suppl 1):s257–s266.
56. Albrektsson T, Isidor F. Consensus report of session IV. In Lang NP, Karring T (eds). *Proceedings of the First European Workshop on Periodontology*. London: Quintessence, 1994:365–369.
57. Wennström JL, Derks J. Is there a need for keratinized mucosa around implants to maintain health and tissue stability?. *Clin Oral Implants Res* 2012;23(suppl 6):136–146.
58. Gobbato L, Avila-Ortiz G, Sohrabi K, Wang CW, Karimbux N. The effect of keratinized mucosa width on peri-implant health: A systematic review. *Int J Oral Maxillofac Implants* 2013;28:1536–1545.
59. Lin GH, Chan HL, Wang HL. The significance of keratinized mucosa on implant health: A systematic review. *J Periodontol* 2013;84(12):1755–1767.
60. Brito C, Tenenbaum HC, Wong BK, Schmitt C, Nogueira-Filho G. Is keratinized mucosa indispensable to maintain peri-implant health? A systematic review of the literature. *J Biomed Mater Res Part B Appl Biomater* 2014;102:643–650.
61. Thoma DS, Mühlemann S, Jung RE. Critical soft-tissue dimensions with dental implants and treatment concepts. *Periodontol* 2000 2014;66:106–118.
62. Chung DM, Oh TJ, Shotwell JL, Misch CE, Wang HL. Significance of keratinized mucosa in maintenance of dental implants with different surfaces. *J Periodontol* 2006;77:1410–1420.
63. Bouri A Jr, Bissada N, Al-Zahrani MS, Faddoul F, Nouneh I. Width of keratinized gingiva and the health status of the supporting tissues around dental implants. *Int J Oral Maxillofac Implants* 2008;23:323–326.
64. Adibrad M, Shahabuei M, Sahabi M. Significance of the width of keratinized mucosa on the health status of the supporting tissue around implants supporting overdentures. *J Oral Implantol* 2009;35:232–237.
65. Rocuzzo M, Grasso G, Dalmasso P. Keratinized mucosa around implants in partially edentulous posterior mandible: 10-year results of a prospective comparative study. *Clin Oral Implants Res* 2016;27:491–496.
66. Boynueğri D, Nemli SK, Kasko YA. Significance of keratinized mucosa around dental implants: A prospective comparative study. *Clin Oral Implants Res* 2013;24:928–933.
67. Krekler G, Kappert HF, Schilli W. Scanning electron microscopic study of the reaction of human bone to a titanium implant. *Int J Oral Surg* 1985;14:447–450.
68. Mericske-Stern R. Clinical evaluation of overdenture restorations supported by osseointegrated titanium implants: a retrospective study. *Int J Oral Maxillofac Implants* 1990;5:375–383.
69. Wennström JL, Bengazi F, Lekholm U. The influence of the masticatory mucosa on the peri-implant soft tissue condition. *Clin Oral Implants Res* 1994;5:1–8.
70. Heckmann SM, Karl M, Wichmann MG, Winter W, Graef F, Taylor TD. Cement fixation and screw retention: Parameters of passive fit. An in vitro study of three-unit implant-supported fixed partial dentures. *Clin Oral Implants Res* 2004;15:466–473.
71. Kim BS, Kim YK, Yun PY, et al. Evaluation of peri-implant tissue response according to the presence of keratinized mucosa. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2009;107:e24–e28.
72. Schrott AR, Jimenez M, Hwang JW, Fiorellini J, Weber HP. Five-year evaluation of the influence of keratinized mucosa on peri-implant soft-tissue health and stability around implants supporting full-arch mandibular fixed prostheses. *Clin Oral Implants Res* 2009;20:1170–1177.
73. Schwarz F, Derks J, Monje A, Wang HL. Peri-implantitis. *J Periodontol* 2018;89(suppl 1):s267–s290.
74. Canullo L, Peñarrocha-Oltra D, Covani U, Botticelli D, Serino G, Peñarrocha M. Clinical and microbiological findings in patients with peri-implantitis: A cross-sectional study. *Clin Oral Implants Res* 2016;27:376–382.
75. Souza AB, Tormena M, Matarazzo F, Araújo MG. The influence of peri-implant keratinized mucosa on brushing discomfort and peri-implant tissue health. *Clin Oral Implants Res* 2016;27:650–655.
76. Ueno D, Nagano T, Watanabe T, Shirakawa S, Yoshima A, Gomi K. Effect of the keratinized mucosa width on the health status of peri-implant and contralateral periodontal tissues: A cross-sectional study. *Implant Dent* 2016;25:796–801.
77. Ladwein C, Schmelzeisen R, Nelson K, Fluegge TV, Fretwurst T. Is the presence of keratinized mucosa associated with periimplant tissue health? A clinical cross-sectional analysis. *Int J Implant Dent* 2015;1:11.
78. Esfahanizadeh N, Daneshparvar N, Motallebi S, Akhondi N, Askarpour F, Davaei S. Do we need keratinized mucosa for a healthy peri-implant soft tissue? *Gen Dent* 2016;64:51–55.
79. Roos-Jansäker AM, Lindahl C, Renvert H, Renvert S. Nine- to fourteen-year follow-up of implant treatment. Part I: Implant loss and associations to various factors. *J Clin Periodontol* 2006;33:283–289.
80. Evan CD, Chen ST. Esthetic outcomes of immediate implant placements. *Clin Oral Implants Res* 2008;19:73–80.
81. Cosyn J, De Bruyn H, Cleymaet R. Soft tissue preservation and pink aesthetics around single immediate implant restorations: A 1-year prospective study. *Clin Implant Dent Relat Res* 2013;15:847–857.
82. Schneider D, Grunder U, Ender A, Hämmerle CH, Jung RE. Volume gain and stability of peri-implant tissue following bone and soft tissue augmentation: 1-year results from a prospective cohort study. *Clin Oral Implants Res* 2011;22:28–37.
83. Thoma DS, Zeltner M, Hilbe M, Hämmerle CH, Hüslér J, Jung RE. Randomized controlled clinical study evaluating effectiveness and safety of a volume-stable collagen matrix compared to autogenous connective tissue grafts for soft tissue augmentation at implant sites. *J Clin Periodontol* 2016;43:874–885.
84. Buyukozdemiraskin S, Berker E, Akincibay H, et al. Necessity of keratinized tissues for dental implants: A clinical, immunological, and radiographic study. *Clin Implant Dent Relat Res* 2015;17:1–12.
85. Bagemez C, Karabuda ZC, Demirel K, Yalcin S. The comparison of acellular dermal matrix allografts with free gingival grafts in the augmentation of peri-implant attached mucosa: A randomised controlled trial. *Eur J Oral Implantol* 2013;6:145–152.
86. Thoma DS, Naenni N, Figuero E, et al. Effects of soft tissue augmentation procedures on peri-implant health or disease: A systematic review and meta-analysis. *Clin Oral Implants Res* 2018;29(suppl 15):32–49.